

## Subject Index

### Volume 63 (1992)

- aggregation, dextran sulfate, liposomes, stearylamine, fusion, fluorescence spectroscopy, freeze fracture electron microscopy, 15
- allene fatty acids, cyclopropene fatty acids, fluorofatty acids, Z11-desaturase, inhibition, *Spodoptera littoralis*, 149
- Amadori compounds, malondialdehyde, lipid oxidation, linolenic acid, non-enzymic browning, 265
- antioxidant,  $\alpha$ -tocopherol, vitamin E, liposomes, spin probe, 69
- antioxidation, flavonoid, linoleic acid, autoxidation, micelle, 37
- argentation chromatography, steroid dienes, dehydrohalogenation, nuclear magnetic resonance spectroscopy, 115
- autoxidation, flavonoid, antioxidation, linoleic acid, micelle, 37
- bilayer modulation, diacylenic phospholipids, liposomes, tubules,  $\beta$ -oxydiacylenic acid, Langmuir film, calorimetry, cooperative unit, optical microscopy, 191
- bilayer, fusion, hexagonal phase  $H_{II}$ , phospholipids, conformation, 251
- bilayer-hexagonal phase transition, tight junctions, non-lamellar lipid phase, fluorescence spectroscopy, calorimetry, 213
- bilayers, dimyristoylphosphatidyl glycerol, melittin, interaction, high pressures, infrared spectra, 139
- C-glucopyranosides, steroidal C-glucosides, C-glucosides, C-glucosidation, steroids, naphthols, 179
- C-glucosidation, steroidal C-glucosides, C-glucopyranosides, C-glucosides, steroids, naphthols, 179
- C-glucosides, steroidal C-glucosides, C-glucopyranosides, C-glucosidation, steroids, naphthols, 179
- $^{13}\text{C}$ -NMR, dihomogamma-linolenic acid, deuterium, lipid, fatty acid, 259
- calorimetry, diacylenic phospholipids, liposomes, tubules, bilayer modulation,  $\beta$ -oxydiacylenic acid, Langmuir film, cooperative unit, optical microscopy, 191
- calorimetry, tight junctions, non-lamellar lipid phase, bilayer-hexagonal phase transition, fluorescence spectroscopy, 213
- cholesterol transfer, phosphatidylserine, ionic strength, pyrene, fluorescence, 55
- cholesterol, phosphatidylserine, phase behaviour, differential scanning calorimetry (DSC), X-ray diffraction, 105
- cholesterol, vesicles, solubilization, octylglucoside, diglycerol hexadecylether, turbidity, gel exclusion HPLC, 1
- cis-unsaturated fatty acid, fatty acid mixture, phase diagram, miscibility, polymorphism, differential scanning calorimetry (DSC), 243
- collapse molecular surface area, diphytanylglycerol ether (archaeol) phospholipids and glycolipids, tetraether (caldarchaeol) phosphoglycolipids and glycolipids, compression isotherms, collapse pressure, limiting molecular area, surface compressional modulus, polar headgroup conformation, 131
- collapse pressure, diphytanylglycerol ether (archaeol) phospholipids and glycolipids, tetraether (caldarchaeol) phosphoglycolipids and glycolipids, compression isotherms, collapse molecular surface area, limiting molecular area, surface compressional modulus, polar headgroup conformation, 131
- compression isotherms, diphytanylglycerol ether (archaeol) phospholipids and glycolipids, tetraether (caldarchaeol) phosphoglycolipids and glycolipids, collapse molecular surface area, collapse pressure, limiting molecular area, surface compressional modulus, polar headgroup conformation, 131
- conformation, bilayer, fusion, hexagonal phase  $H_{II}$ , phospholipids, 251
- conjugated diacylenic fatty acids, mass spectrometry, piconyl esters, 65
- cooperative unit, diacylenic phospholipids, liposomes, tubules, bilayer modulation,  $\beta$ -oxydiacylenic acid, Langmuir film, calorimetry, optical microscopy, 191
- cyclopropene fatty acids, allene fatty acids, fluorofatty acids, Z11-desaturase, inhibition, *Spodoptera littoralis*, 149
- dehydrohalogenation, steroid dienes, argentation chromatography, nuclear magnetic resonance spectroscopy, 115
- deuterium, dihomogamma-linolenic acid,  $^{13}\text{C}$ -NMR, lipid, fatty acid, 259
- dextran sulfate, liposomes, stearylamine, aggregation, fusion, fluorescence spectroscopy, freeze fracture electron microscopy, 15
- diacylenic phospholipids, liposomes, tubules, bilayer modulation,  $\beta$ -oxydiacylenic acid, Langmuir film, calorimetry, cooperative unit, optical microscopy, 191
- differential scanning calorimetry (DSC), cis-unsaturated fatty acid, fatty acid mixture, phase diagram, miscibility, polymorphism, 243
- differential scanning calorimetry (DSC), phosphatidylserine, cholesterol, phase behaviour, X-ray diffraction, 105
- differential scanning calorimetry, phosphatidic acids, phase diagram, hydrated PA mixture, unhydrated PA mixture, miscibility, 203
- diglycerol hexadecylether, vesicles, solubilization, octylglucoside, cholesterol, turbidity, gel exclusion HPLC, 1
- dihomogamma-linolenic acid, deuterium,  $^{13}\text{C}$ -NMR, lipid, fatty acid, 259
- dimyristoylphosphatidyl glycerol, bilayers, melittin, interaction, high pressures, infrared spectra, 139

- diosmin, flavonoids, model membranes, elasticity modulus, quasi-elastic light scattering, 169
- diphytanylglycerol ether (archaeol) phospholipids and glycolipids, tetraether (caldarchaeol) phosphoglycolipids and glycolipids, compression isotherms, collapse molecular surface area, collapse pressure, limiting molecular area, surface compressional modulus, polar headgroup conformation, 131
- elasticity modulus, flavonoids, diosmin, model membranes, quasi-elastic light scattering, 169
- electron paramagnetic resonance (EPR), low-density lipoprotein (LDL) modification, LDL, stabilization of structure, Mn(II) binding, 159
- epithelial cells, tight junctions, resistance, non-lamellar lipid phase, 223
- ether lipid, phospholipid analog, monolayer, surface pressure, 27
- fatty acid mixture, *cis*-unsaturated fatty acid, phase diagram, miscibility, polymorphism, differential scanning calorimetry (DSC), 243
- fatty acid, dihomogamma-linolenic acid, deuterium,  $^{13}\text{C}$ -NMR, lipid, 259
- flavonoid, antioxidation, linoleic acid, autoxidation, micelle, 37
- flavonoids, diosmin, model membranes, elasticity modulus, quasi-elastic light scattering, 169
- fluorescence spectroscopy, dextran sulfate, liposomes, stearylamine, aggregation, fusion, freeze fracture electron microscopy, 15
- fluorescence spectroscopy, tight junctions, non-lamellar lipid phase, bilayer-hexagonal phase transition, calorimetry, 213
- fluorescence, cholesterol transfer, phosphatidylserine, ionic strength, pyrene, 55
- fluorofatty acids, cyclopropene fatty acids, allene fatty acids, Z11-desaturase, inhibition, *Spodoptera littoralis*, 149
- Fourier transform infrared spectrometry, pulmonary surfactant, pulmonary surfactant protein B (SP-B), pulmonary surfactant protein C, monomeric form (SP-C), secondary structure, 91
- freeze fracture electron microscopy, dextran sulfate, liposomes, stearylamine, aggregation, fusion, fluorescence spectroscopy, 15
- fusion, bilayer, hexagonal phase  $\text{H}_{\text{II}}$ , phospholipids, conformation, 251
- fusion, dextran sulfate, liposomes, stearylamine, aggregation, fluorescence spectroscopy, freeze fracture electron microscopy, 15
- gel exclusion HPLC, vesicles, solubilization, octylglucoside, cholesterol, diglycerol hexadecylether, turbidity, 1
- $^1\text{H}$ - and  $^{13}\text{C}$ -NMR, 15-oxygenated sterols, inhibitors of sterol synthesis, 77
- $^2\text{H}$ -NMR, plant sterol, soybean phosphatidylcholine, oriented bilayers, 235
- hexagonal phase  $\text{H}_{\text{II}}$ , bilayer, fusion, phospholipids, conformation, 251
- high pressures, dimyristoylphosphatidyl glycerol, bilayers, melittin, interaction, infrared spectra, 139
- hydrated PA mixture, phosphatidic acids, phase diagram, unhydrated PA mixture, miscibility, differential scanning calorimetry, 203
- infrared spectra, dimyristoylphosphatidyl glycerol, bilayers, melittin, interaction, high pressures, 139
- inhibition, cyclopropene fatty acids, allene fatty acids, fluorofatty acids, Z11-desaturase, *Spodoptera littoralis*, 149
- inhibitors of sterol synthesis, 15-oxygenated sterols,  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR, 77
- interaction, dimyristoylphosphatidyl glycerol, bilayers, melittin, high pressures, infrared spectra, 139
- ionic strength, cholesterol transfer, phosphatidylserine, pyrene, fluorescence, 55
- Langmuir film, diacylenic phospholipids, liposomes, tubules, bilayer modulation,  $\beta$ -oxydiacylenic acid, calorimetry, cooperative unit, optical microscopy, 191
- LDL, stabilization of structure, electron paramagnetic resonance (EPR), low-density lipoprotein (LDL) modification, Mn(II) binding, 159
- light scattering, polymerizable lipid, phosphocholine, lipid microstructures, phase properties, 47
- limiting molecular area, diphytanylglycerol ether (archaeol) phospholipids and glycolipids, tetraether (caldarchaeol) phosphoglycolipids and glycolipids, compression isotherms, collapse molecular surface area, collapse pressure, surface compressional modulus, polar headgroup conformation, 131
- linoleic acid, flavonoid, antioxidation, autoxidation, micelle, 37
- linolenic acid, Amadori compounds, malondialdehyde, lipid oxidation, non-enzymic browning, 265
- lipid microstructures, polymerizable lipid, phosphocholine, phase properties, light scattering, 47
- lipid oxidation, Amadori compounds, malondialdehyde, linolenic acid, non-enzymic browning, 265
- lipid, dihomogamma-linolenic acid, deuterium,  $^{13}\text{C}$ -NMR, fatty acid, 259
- liposomes,  $\alpha$ -tocopherol, vitamin E, spin probe, antioxidant, 69
- liposomes, dextran sulfate, stearylamine, aggregation, fusion, fluorescence spectroscopy, freeze fracture electron microscopy, 15
- liposomes, diacylenic phospholipids, tubules, bilayer modulation,  $\beta$ -oxydiacylenic acid, Langmuir film, calorimetry, cooperative unit, optical microscopy, 191
- low-density lipoprotein (LDL) modification, electron paramagnetic resonance (EPR), LDL, stabilization of structure, Mn(II) binding, 159
- malondialdehyde, Amadori compounds, lipid oxidation, linolenic acid, non-enzymic browning, 265
- mass spectrometry, conjugated diacylenic fatty acids, picolinyl esters, 65
- mass spectrometry, mycolic acids, thermospray, 41
- melittin, dimyristoylphosphatidyl glycerol, bilayers, interaction, high pressures, infrared spectra, 139

- micelle, flavonoid, antioxidation, linoleic acid, autoxidation, 37
- miscibility, *cis*-unsaturated fatty acid, fatty acid mixture, phase diagram, polymorphism, differential scanning calorimetry (DSC), 243
- miscibility, phosphatidic acids, phase diagram, hydrated PA mixture, unhydrated PA mixture, differential scanning calorimetry, 203
- Mn(II) binding, electron paramagnetic resonance (EPR), low density lipoprotein (LDL) modification, LDL, stabilization of structure, 159
- model membranes, flavonoids, diosmin, elasticity modulus, quasi-elastic light scattering, 169
- monolayer, phospholipid analog, ether lipid, surface pressure, 27
- mycolic acids, mass spectrometry, thermospray, 41
- naphthols, steroidal C-glucosides, C-glucopyranosides, C-glucosides, C-glucosidation, steroids, 179
- non-enzymic browning, Amadori compounds, malondialdehyde, lipid oxidation, linolenic acid, 265
- non-lamellar lipid phase, epithelial cells, tight junctions, resistance, 223
- non-lamellar lipid phase, tight junctions, bilayer-hexagonal phase transition, fluorescence spectroscopy, calorimetry, 213
- nuclear magnetic resonance spectroscopy, steroid dienes, dehydrohalogenation, argentation chromatography, 115
- octylglucoside, vesicles, solubilization, cholesterol, diglycerol hexadecylether, turbidity, gel exclusion HPLC, 1
- optical microscopy, diacylenic phospholipids, liposomes, tubules, bilayer modulation,  $\beta$ -oxydiacylenic acid, Langmuir film, calorimetry, cooperative unit, 191
- oriented bilayers,  $^2\text{H}$ -NMR, plant sterol, soybean phosphatidylcholine, 235
- $\beta$ -oxydiacylenic acid, diacylenic phospholipids, liposomes, tubules, bilayer modulation, Langmuir film, calorimetry, cooperative unit, optical microscopy, 191
- 15-oxygenated sterols, inhibitors of sterol synthesis,  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR, 77
- 15-oxygenated sterols, steroids, synthesis, 23
- phase behaviour, phosphatidylserine, cholesterol, differential scanning calorimetry (DSC), X-ray diffraction, 105
- phase diagram, *cis*-unsaturated fatty acid, fatty acid mixture, miscibility, polymorphism, differential scanning calorimetry (DSC), 243
- phase diagram, phosphatidic acids, hydrated PA mixture, unhydrated PA mixture, miscibility, differential scanning calorimetry, 203
- phase properties, polymerizable lipid, phosphocholine, lipid microstructures, light scattering, 47
- phosphatidic acids, phase diagram, hydrated PA mixture, unhydrated PA mixture, miscibility, differential scanning calorimetry, 203
- phosphatidylserine, cholesterol transfer, ionic strength, pyrene, fluorescence, 55
- phosphatidylserine, cholesterol, phase behaviour, differential scanning calorimetry (DSC), X-ray diffraction, 105
- phosphocholine, polymerizable lipid, lipid microstructures, phase properties, light scattering, 47
- phospholipid analog, ether lipid, monolayer, surface pressure, 27
- phospholipids, bilayer, fusion, hexagonal phase  $\text{H}_{\text{II}}$ , conformation, 251
- picolinyl esters, conjugated diacylenic fatty acids, mass spectrometry, 65
- plant sterol,  $^2\text{H}$ -NMR, soybean phosphatidylcholine, oriented bilayers, 235
- polar headgroup conformation, diphytanylglycerol ether (archaeol) phospholipids and glycolipids, tetraether (caldarchaeol) phosphoglycolipids and glycolipids, compression isotherms, collapse molecular surface area, collapse pressure, limiting molecular area, surface compressional modulus, 131
- polymerizable lipid, phosphocholine, lipid microstructures, phase properties, light scattering, 47
- polymorphism, *cis*-unsaturated fatty acid, fatty acid mixture, phase diagram, miscibility, differential scanning calorimetry (DSC), 243
- pulmonary surfactant protein B (SP-B), pulmonary surfactant, pulmonary surfactant protein C, monomeric form (SP-C), Fourier transform infrared spectrometry, secondary structure, 91
- pulmonary surfactant protein C, monomeric form (SP-C), pulmonary surfactant, pulmonary surfactant protein B (SP-B), Fourier transform infrared spectrometry, secondary structure, 91
- pulmonary surfactant, pulmonary surfactant protein B (SP-B), pulmonary surfactant protein C, monomeric form (SP-C), Fourier transform infrared spectrometry, secondary structure, 91
- pyrene, cholesterol transfer, phosphatidylserine, ionic strength, fluorescence, 55
- quasi-elastic light scattering, flavonoids, diosmin, model membranes, elasticity modulus, 169
- resistance, epithelial cells, tight junctions, non-lamellar lipid phase, 223
- secondary structure, pulmonary surfactant, pulmonary surfactant protein B (SP-B), pulmonary surfactant protein C, monomeric form (SP-C), Fourier transform infrared spectrometry, 91
- solubilization, vesicles, octylglucoside, cholesterol, diglycerol hexadecylether, turbidity, gel exclusion HPLC, 1
- soybean phosphatidylcholine,  $^2\text{H}$ -NMR, plant sterol, oriented bilayers, 235
- spin probe,  $\alpha$ -tocopherol, vitamin E, liposomes, antioxidant, 69
- Spodoptera littoralis*, cyclopropane fatty acids, allene fatty acids, fluorofatty acids, Z11-desaturase, inhibition, 149
- stearylamine, dextran sulfate, liposomes, aggregation, fusion, fluorescence spectroscopy, freeze fracture electron microscopy, 15
- sterol dienes, dehydrohalogenation, argentation chromatography, nuclear magnetic resonance spectroscopy, 115

steroidal C-glucosides, C-glucopyranosides, C-glucosides, C-glucosidation, steroids, naphthols, 179

steroids, 15-oxygenated sterols, synthesis, 23

steroids, steroidal C-glucosides, C-glucopyranosides, C-glucosides, C-glucosidation, naphthols, 179

surface compressional modulus, diphytanylglycerol ether (archaeol) phospholipids and glycolipids, tetraether (caldarchaeol) phosphoglycolipids and glycolipids, compression isotherms, collapse molecular surface area, collapse pressure, limiting molecular area, polar headgroup conformation, 131

surface pressure, phospholipid analog, ether lipid, monolayer, 27

synthesis, steroids, 15-oxygenated sterols, 23

tetraether (caldarchaeol) phosphoglycolipids and glycolipids, diphytanylglycerol ether (archaeol) phospholipids and glycolipids, compression isotherms, collapse molecular surface area, collapse pressure, limiting molecular area, surface compressional modulus, polar headgroup conformation, 131

thermospray, mycolic acids, mass spectrometry, 41

tight junctions, epithelial cells, resistance, non-lamellar lipid phase, 223

tight junctions, non-lamellar lipid phase, bilayer-hexagonal phase transition, fluorescence spectroscopy, calorimetry, 213

$\alpha$ -tocopherol, vitamin E, liposomes, spin probe, antioxidant, 69

tubules, diacylenic phospholipids, liposomes, bilayer modulation,  $\beta$ -oxydiacetylenic acid, Langmuir film, calorimetry, cooperative unit, optical microscopy, 191

turbidity, vesicles, solubilization, octylglucoside, cholesterol, diglycerol hexadecylether, gel exclusion HPLC, 1

unhydrated PA mixture, phosphatidic acids, phase diagram, hydrated PA mixture, miscibility, differential scanning calorimetry, 203

vesicles, solubilization, octylglucoside, cholesterol, diglycerol hexadecylether, turbidity, gel exclusion HPLC, 1

vitamin E,  $\alpha$ -tocopherol, liposomes, spin probe, antioxidant, 69

X-ray diffraction, phosphatidylserine, cholesterol, phase behaviour, differential scanning calorimetry (DSC), 105

Z11-desaturase, cyclopropane fatty acids, allene fatty acids, fluorofatty acids, inhibition, *Spodoptera littoralis*, 149

## Author Index

### Volume 63 (1992)

Absolom, D.R.	91	Hermosín, I.	265
Ahmed, M.K.	139	Hiramatsu, N.	243
Allevi, P.	179	Horváth, J.	23
Anastasia, M.	179		
Arnold, K.	15	Ikebata, W.	69
Arsequell, G.	149	Inoue, T.	203, 243
Azize, B.	169	Ioneda, T.	41
Baatz, J.E.	91	Jürgens, G.	159
Bach, D.	105		
Bakos, T.	23	Kahn, B.	47
Baral-Tosh, S.	47	Kates, M.	131
Baxter, C.	91	Kinnunen, P.K.J.	251
Beaman, B.L.	41	Krajewski-Bertrand, M-A.	235
Borochoy, N.	105	Kumadaki, I.	69
Brechany, E.Y.	65		
Brnjas-Kraljević, J.	159	Lebeau, L.	27
		Ledl, F.	265
Camps, F.	149	Lesieur, S.	1
Cao, A.	169	Lewin, G.	169
Chau, S.H.	65	Li, Q-T.	55
Chen, Z-Y.	259	Lie Ken Jie, M.S.F.	65
Cheung, Y.K.	65		
Chifu, E.	131	Markowitz, M.A.	191
Choma, C.T.	139	Mädefessel, C.	223
Christie, W.W.	65	Mendy, F.	259
Ciuffreda, P.	179	Mésini, P.	27
Cunnane, S.C.	259	Milon, A.	235
		Mioskowski, C.	27
Descomps, B.	259	Mocanu, A.	131
		Motoda, I.	243
Epand, R.M.	105		
		Ohki, S.	15
Fabriás, G.	149	Ollivon, M.	1
Fukuzawa, K.	69	Oudet, P.	27
Galla, H-J.	213, 223	Perret, G.	169
Gómez-Sánchez, A.	265	Pifat, G.	159
Gosalbo, L.	149	Pinkerton, F.D.	77
		Post, A.	213, 223
Haag, B.	223		
Handjani-Vila, R-M.	1	Quinn, P.J.	131
Hantz-Brachet, E.	169	Ratna, B.R.	47
Hartmann, M-A.	235	Richter, W.	15
Haycock, P.R.	259	Rudolph, A.S.	47
Hein, M.	213, 223	Ruecker, K.E.	77
Herak, J.N.	159		
Herak-Kramberger, C.M.	159	Sakanaka, T.	69

Sanvito, A.M.	179	Tomoia-Cotisel, M.	131
Sato, K.	243	Tsao, L.-I.	191
Sawyer, W.H.	55		
Scala, A.	179	Urano, S.	69
Schnur, J.M.	47		
Schroepfer Jr., G.J.	77, 115	Vincze, I.	23
Senisterra, G.	105		
Seras, M.	1	Wachtel, E.	105
Shibata, A.	69	Wang, P.-F.	37
Shimozawa, R.	203	Whitsett, J.A.	91
Siddiqui, A.U.	77, 115	Williams, S.C.R.	259
Singh, A.	191	Wilson, W.K.	77, 115
Smyth, K.L.	91	Wong, P.T.T.	139
Suzuki, M.	243		
Swaminathan, S.	115	Zheng, R.-L.	37
		Zsako, J.	131
Taillandier, E.	169	Zschörnig, O.	15
Tasaka, T.	203	CPL2138	
Teichmann, K.	223		



